

A ratchet mechanism for amplification in low-frequency mammalian hearing

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The sensitivity and frequency selectivity of hearing result from tuned amplification by an active process in the mechanoreceptive hair cells. In most vertebrates, the active process stems from the active motility of hair bundles. The mammalian cochlea exhibits an additional form of mechanical activity termed electromotility: its outer hair cells (OHCs) change length upon electrical stimulation. The relative contributions of these two mechanisms to the active process in the mammalian inner ear is the subject of intense current debate. Here, we show that active hair-bundle motility and electromotility can together implement an efficient mechanism for amplification that functions like a ratchet: Sound-evoked forces, acting on the basilar membrane, are transmitted to the hair bundles, whereas electromotility decouples active hair-bundle forces from the basilar membrane. This unidirectional coupling can extend the hearing range well below the resonant frequency of the basilar membrane. It thereby provides a concept for low-frequency hearing that accounts for a variety of unexplained experimental observations from the cochlear apex, including the shape and phase behavior of apical tuning curves, their lack of significant nonlinearities, and the shape changes of threshold tuning curves of auditory-nerve fibers along the cochlea. The ratchet mechanism constitutes a general design principle for implementing mechanical amplification in engineering applications.

auditory system | cochlea | hair cell

The mammalian cochlea acts as a frequency analyzer in which high frequencies are detected at the organ's base and low frequencies at more apical positions. This frequency mapping is thought to be achieved by a position-dependent resonance of the elastic basilar membrane separating two fluid-filled compartments (Fig. 1A) (1–3). When sound evokes a pressure wave that displaces the basilar membrane, the resultant traveling wave gradually increases in amplitude as it progresses to the position where the basilar membrane's resonant frequency coincides with that of the stimulus. Aided by mechanical energy provided by the active process, the wave peaks at a characteristic place slightly before the resonant position and then declines sharply (Fig. 1B). This mechanism is termed critical-layer absorption (1), for a wave cannot travel beyond its characteristic position on the basilar membrane but peaks and dissipates most of its energy there. The mechanism displays scale invariance: different stimulation frequencies induce traveling waves that display a common, strongly asymmetric form upon rescaling of the amplitude and spatial coordinate (4, 5).

Two important aspects of the cochlea's mechanics remain problematical. First, the basilar membrane's resonant frequency apparently cannot span the entire range of audible frequencies. Experimental measurements of basilar-membrane stiffness suggest that high-frequency resonances are feasible but low-frequency ones are inaccessible (6). This result accords with the analysis of threshold tuning curves for auditory-nerve fibers (7–9) and measurements of basilar-membrane displacement (8, 10–12), both of which indicate that a peaked traveling wave occurs for high-frequency stimulation but not for low-frequency stimulation. Indeed, threshold tuning curves of auditory-nerve fibers (7–9) show that high-frequency curves are scale-invariant and possess

a sharp cutoff at frequencies above their characteristic frequencies, reflecting the mechanism of critical-layer absorption. However, low-frequency curves lack this sharp cutoff and, in contradiction of the expectation for critical-layer absorption, are instead characterized by an approximately symmetric shape around their characteristic frequencies. Recent experiments in the chinchilla have shown that the shape change between the tuning curves of high- and low-frequency fibers occurs between two crossover frequencies of about 5 kHz and 1.5 kHz (8, 9). Measurements of basilar-membrane displacement yield similar conclusions. Experiments from the cochlear base confirm the existence at high frequencies of peaked traveling waves that result in strongly asymmetric tuning curves and a pronounced nonlinearity at the characteristic frequencies (8, 12–14). Apical measurements of basilar-membrane displacement, however, produce symmetric tuning curves that lack a sharp cutoff for frequencies higher than the characteristic frequency (10, 11, 15). Moreover, only small nonlinearity has been measured (10, 15, 16), raising the question whether amplification occurs at the apex. All available experimental results therefore indicate that low-frequency hearing does not function through critical-layer absorption but must rely on another mechanism to achieve frequency selectivity.

The second key uncertainty is the nature of the active process that operates in the mammalian cochlea. In addition to amplifying weak signals, the active process produces increased frequency selectivity, compressive nonlinearity, and spontaneous otoacoustic emission. In the mammalian cochlea, amplification is provided by specialized OHCs located in the organ of Corti along the basilar membrane (Fig. 24). Each OHC displays two forms of motility. Like those in the hearing organs of other vertebrates (17, 18), the hair bundle of an OHC can produce mechanical force (17–22). But an OHC also exhibits electromotility: when its membrane potential changes as a result of sound-evoked hair-bundle deflection, the entire cell undergoes a length change owing to conformational rearrangement of the membrane protein prestin (23–25). Direct measurements of the respective roles of the two forms of motility are complicated, for it is difficult to determine the micromechanical responses of the organ of Corti while the cochlea remains intact.

Here we have taken a theoretical approach to investigate a possible mechanism for amplification by the synergistic interplay of active hair-bundle motility and electromotility. We show that they can operate together to achieve low-frequency selectivity in the absence of basilar-membrane resonance and critical-layer absorption. The model advances theoretical understanding of cochlear mechanics in three ways: it reproduces previous theoretical results insofar as they coincide with experiments; it accounts

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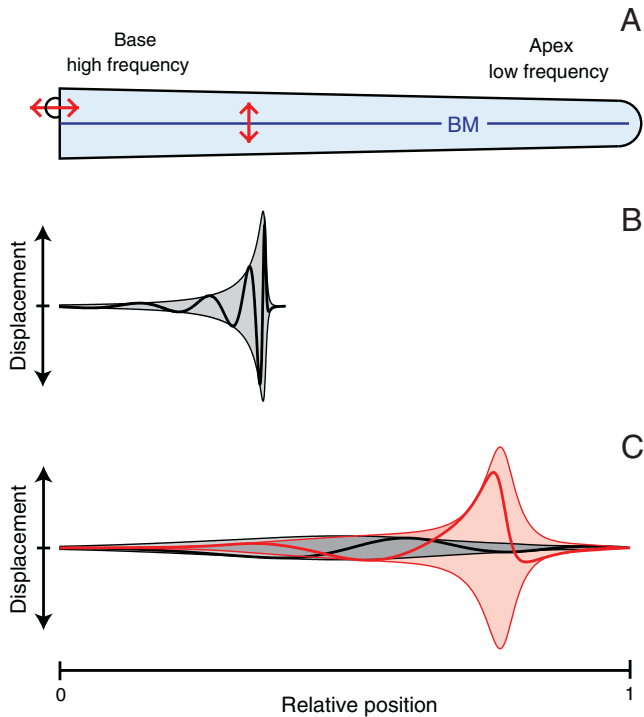


Fig. 1. Principles of cochlear mechanics. (A) In a schematic diagram of the mammalian cochlea, the basilar membrane (BM) is displaced by sound stimuli acting on the stapes (Upper Left). (B) In the classical theory of cochlear mechanics, sound evokes a pressure wave that causes a longitudinal traveling wave of basilar-membrane displacement (Thick Line). The motion of the basilar membrane and the displacements of the associated hair bundles are approximately equal. As the wave approaches the position where its frequency matches the basilar membrane's resonant frequency, the wave's amplitude (Thin Line) increases and its wavelength and velocity decline. The wave peaks at a characteristic place slightly before the resonant position and then declines sharply, yielding a strongly asymmetric envelope of the traveling wave (Shading). Experiments confirm this behavior in the basal, high-frequency part of the cochlea. (C) We propose an alternative theory for the cochlea's mechanics at low frequencies. The basilar membrane near the cochlear apex does not resonate, but the traveling wave on the basilar membrane propagates along the entire cochlea without a strong variation in amplitude, wavelength, and velocity (Black). However, the interplay of electromotility and active hair-bundle motility fosters an independent resonance of the complex formed by the hair bundles, reticular lamina, and tectorial membrane. The hair-bundle displacement (Red) at the characteristic place can therefore exhibit an approximately symmetric peak, exceeding basilar membrane motion by orders of magnitude.

naturally for a variety of unexplained findings from the cochlear apex; and it makes robust, experimentally testable predictions.

Results

The Ratchet Mechanism. Consider a transverse element of the cochlear partition comprising the basilar membrane, an inner hair cell (IHC) and three OHCs, and the overlying tectorial membrane (Fig. 2A). When a sound-evoked force displaces the basilar membrane upward, the resultant shearing motion between the tectorial membrane and the top of the OHCs, or reticular lamina, deflects the hair bundles in the positive direction. Active hair-bundle motility in the OHCs increases the amplitude of deflection. Without electromotility, this additional displacement would couple back to the basilar membrane and augment the movement there. If electromotility is adjusted such that the OHCs elongate just as much as the tectorial membrane and reticular lamina move upward, however, the basilar membrane does not experience the active force and thus undergoes no additional displacement (Movie S1).

A mathematical formulation of this amplification mechanism clarifies its operation. Consider a two-mass model in which the

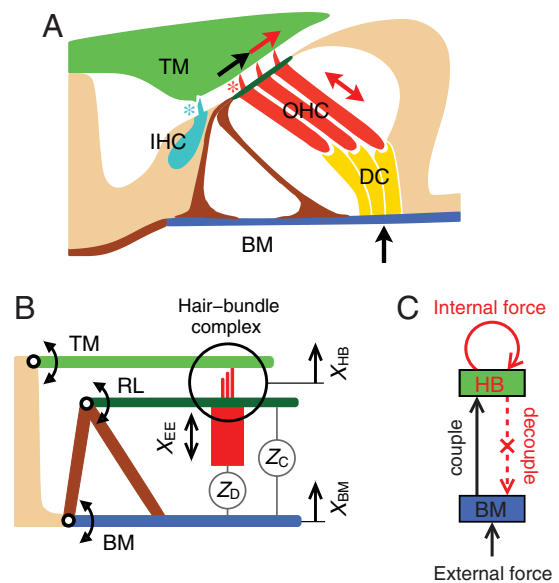


Fig. 2. The ratchet mechanism. (A) The organ of Corti rests upon the basilar membrane (BM). Three OHCs are connected to Deiters' cells (DC) that together couple the basilar membrane to the reticular lamina (Dark Green, top of the OHCs) and through the hair bundles to the overlying tectorial membrane (TM). Sound-evoked external forces (Black Arrow) displace the basilar membrane, here upwards, and produce shearing (Black Arrow) of the hair bundles of OHCs (Red Asterisk) and the inner hair cell (IHC) (Cyan Asterisk). Two forms of motility underlie the active process: active hair-bundle motility (Single-Headed Red Arrow) and membrane-based electromotility (Double-Headed Red Arrow). (B) The two fundamental degrees of freedom are the basilar-membrane displacement X_{BM} and the displacement X_{HB} of the hair-bundle complex (circle) that comprises the hair bundles, reticular lamina (RL), and tectorial-membrane. Coupling stems from the impedance Z_D of the combined OHCs and Deiters' cells as well as the impedance Z_C of the remaining organ of Corti. (C) In the ratchet mechanism, displacements of the basilar membrane caused by external forces are communicated to the hair-bundle complex. Internal forces (Red Arrow) in the hair bundles increase the shearing motion that decouples from basilar-membrane displacement through appropriate length changes of the OHCs (Dotted Red Arrow). For an animated representation of the model, see Movie S1.

two degrees of freedom are the motion of the basilar membrane, X_{BM} , and that of the complex formed by the hair bundles, reticular lamina, and tectorial membrane, X_{HB} (Fig. 2B). A sound stimulus of frequency f produces an oscillating external force $F_{ext}(t) = \tilde{F}_{ext}e^{2\pi ift} + c.c.$ acting on the basilar membrane, in which $c.c.$ denotes the complex conjugate. The evoked oscillations of the basilar membrane as well as the hair-bundle complex occur predominantly at the same frequency, $X_{BM} = \tilde{X}_{BM}e^{2\pi ift} + c.c.$ and $X_{HB} = \tilde{X}_{HB}e^{2\pi ift} + c.c.$. Internal forces F_{int} arise within hair bundles, where they provide negative damping and introduce nonlinearities (SI Text). The cell bodies of OHCs can be described as piezoelectric elements (26). For small, physiologically relevant motions their electrically evoked displacement $X_{EE} = \tilde{X}_{EE}e^{2\pi ift} + c.c.$ is proportional to the hair-bundle displacement that triggers changes in the membrane potential, so $\tilde{X}_{EE} = -\alpha\tilde{X}_{HB} + c.c.$ with a complex mechanomotility coefficient α (27). The hair-bundle and basilar-membrane displacements then depend linearly on the forces:

$$A \begin{pmatrix} \tilde{X}_{HB} \\ \tilde{X}_{BM} \end{pmatrix} = \frac{1}{2\pi if} \begin{pmatrix} \tilde{F}_{int} \\ \tilde{F}_{ext} \end{pmatrix}. \quad [1]$$

The matrix A contains the impedances Z_{HB} and Z_{BM} of the hair-bundle complex and the basilar membrane as well as the coupling impedances Z_D and Z_C (Fig. 2B):

$$A = \begin{pmatrix} Z_{HB} + (1 + \alpha)Z_D + Z_C & -Z_D - Z_C \\ -(1 + \alpha)Z_D - Z_C & Z_{BM} + Z_D + Z_C \end{pmatrix}. \quad [2]$$

A key feature of this relation is that the matrix element A_{21} that describes the coupling of the internal force to the basilar membrane vanishes at a critical value $\alpha_* \equiv -1 - Z_C/Z_D$. At the same time, the coupling of the external force to the hair bundle that is represented by element A_{12} remains nonzero. For the critical value α_* the displacements become

$$\begin{aligned} \tilde{X}_{HB} &= \frac{1}{2\pi i f Z_{HB}} \tilde{F}_{int} + \frac{Z_D + Z_C}{2\pi i f Z_{HB}(Z_{BM} + Z_D + Z_C)} \tilde{F}_{ext}, \\ \tilde{X}_{BM} &= \frac{1}{2\pi i f (Z_{BM} + Z_D + Z_C)} \tilde{F}_{ext}. \end{aligned} \quad [3]$$

The hair-bundle displacement depends on both the internal and external forces, whereas the basilar membrane experiences only the external force. In other words, the sound-evoked external force on the basilar membrane is transmitted to the hair bundle, but the internal force from active hair-bundle motility does not feed back onto the motion of the basilar membrane (Fig. 2C). This symmetry-breaking mode has the characteristics of a ratchet in the sense that information flows unidirectionally: Information applied as a force against the basilar membrane can be detected in the form of hair-bundle displacement, but force acting on the hair bundle cannot be detected at the basilar membrane. The basilar-membrane displacement equals that occurring when the hair-bundle complex is fixed at $\tilde{X}_{HB} = 0$ and electromotility is absent ($\alpha = 0$). The ratchet mechanism, therefore, resembles an ideal operational amplifier that neither feeds back on nor draws energy from the input (28).

The Mechanomotility Coefficient α . The magnitude and phase of the critical value α_* depend on the coupling impedances Z_C and Z_D . Under realistic assumptions $|Z_C|$ is similar to or smaller than $|Z_D|$ and $|\alpha_*|$ is therefore near unity. Such a value for the mechanomotility coefficient α has been measured experimentally for apical OHCs (27). The phase of α is controlled by the complex network of ion channels that regulates the membrane potential depending on the hair-bundle displacement (27, 29). Although experiments on the phase of α are not available, theoretical considerations confirm that OHCs have sufficient flexibility through ion-channel regulation to adjust the phase of α to a variety of values and, in particular, to the phase required for α_* (SI Text).

A Concept for Low-Frequency Hearing. As its most important characteristic, the ratchet mechanism can explain frequency selectivity near the cochlear apex in the absence of a peaked traveling wave. The resonant frequency of the hair-bundle complex is determined solely by the impedance Z_{HB} and the internal forces, and is therefore defined by the properties of the hair bundles and tectorial membrane (3) (3031–32). In particular, hair-bundle resonance can occur in the absence of basilar-membrane resonance. The ratchet mechanism therefore permits the resonant frequency of the hair bundles to follow a logarithmic law along the cochlea, whereas the resonant frequency of the basilar membrane remains significantly greater near the apex (Fig. 3B) (6).

Further evidence points to the occurrence of the ratchet mechanism at the cochlear apex but not at the base. First, the basilar membrane at the base is narrow and presumably tuned to the characteristic frequencies at which auditory-nerve fibers are most sensitive (Fig. 3B). Amplification of basilar-membrane motion there is feasible and need not be avoided. Indeed, experiments have demonstrated a strong compressive nonlinearity of basilar-membrane motion at high frequencies. However, the apex exhibits a wider basilar membrane that experiences stronger viscous forces and whose resonant frequencies deviate from the charac-

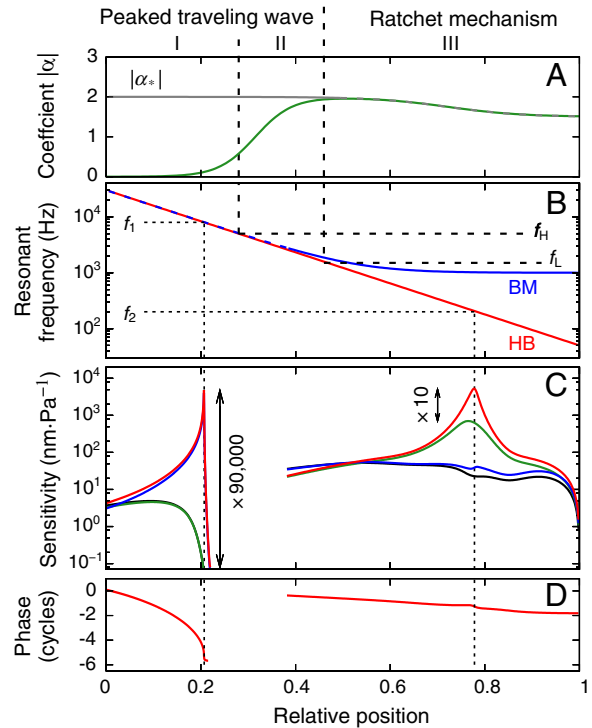


Fig. 3. Cochlear model. (A) The ratchet mechanism operates when the mechanomotility coefficient α (Green) coincides with the critical value α_* (gray). Although electromotility is negligible to the basal side of f_H , it underlies the ratchet mechanism apical to the position of f_L . (B) The resonant frequency of the hair-bundle complex (HB, Red) agrees with that of the basilar membrane (BM, Blue) only for frequencies above f_L . (C) A high-frequency sound stimulus ($f_1 = 8$ kHz) induces a traveling wave that peaks in the basal region. The displacements of the hair bundles (Red) coincide with that of the basilar membrane (Blue). Elimination of active hair-bundle motility decreases the sensitivity by a factor of 90,000 (Green, hair bundles; Black, basilar membrane) indicative of a strong nonlinearity. A low-frequency stimulus ($f_2 = 200$ Hz) triggers a traveling wave that does not peak on the basilar membrane, but the hair-bundle displacement exhibits a resonance enabled by the ratchet mechanism (same color code). Without active hair-bundle motion the hair-bundle displacement decreases by a factor of only 10, indicative of a weak nonlinearity. (D) The phase of the basilar-membrane displacement for f_1 has a strongly increasing slope near the resonant position and thus shows a wave traveling to the resonant position but not beyond. For f_2 the slope of the phase remains almost constant, corresponding to a wave traveling beyond the characteristic place.

teristic frequencies (6). Amplification of basilar-membrane motion near the apex would therefore be highly inefficient. The absence of a strong, compressive nonlinearity in experimental measurements of basilar-membrane displacement in the apical region confirms the lack of basilar-membrane amplification there. Next, the membrane time constant of OHCs restricts electromotility's ability to work on a cycle-by-cycle basis to frequencies below a few kilohertz (33, 34), disabling the ratchet mechanism for higher frequencies. It is noteworthy that the ion channels of apical OHCs differ significantly from those of basal OHCs (35) and that apical OHCs are considerably longer than basal ones and can accordingly produce greater length changes (25).

A Unified Model for Cochlear Mechanics. We have quantified these considerations in a one-dimensional model of the cochlea (SI Text). Studies of threshold tuning curves of auditory-nerve fibers in the chinchilla have delineated three distinct regimes that are separated by two crossover frequencies, a high frequency $f_H \approx 5$ kHz, and a low frequency $f_L \approx 1.5$ kHz (Fig. 3) (9). In our model for the basal regime I, above f_H , electromotility is

negligible owing to the membrane time constant; each segment of the basilar-membrane is tuned to its characteristic frequency. In the transitional regime II, between f_H and f_L , unidirectional coupling provided by electromotility occurs, but every segment of the basilar-membrane still resonates at its characteristic frequency. In the apical regime III, below f_L , electromotility combines with active hair-bundle motility to implement the ratchet mechanism for amplification. The resonant frequency of the basilar-membrane is therefore of minor importance and remains approximately constant at a value significantly above the range of characteristic frequencies.

In accordance with the theory of critical-layer absorption (1), in regime I the traveling-wave produced by a pure-tone stimulus peaks at the characteristic place and sharply decreases beyond that point (Fig. 3C). The characteristic behavior of critical-layer absorption—decrease of the traveling-wave’s speed and wavelength near the characteristic place—appears in the phase behavior as an increase of the slope (Fig. 3D). Active hair-bundle motility amplifies the peak displacement. A strong compressive nonlinearity arises because amplification by active hair-bundle motility both increases the hair-bundle and basilar-membrane motion per unit pressure difference and enhances the amplitude of the pressure wave itself (Fig. 3C). The combination of the two effects yields a compressive nonlinearity that extends over a significantly broader range of sound intensities than the nonlinearity in hair-bundle motion itself.

In regime II, electromotility influences the micromechanics and causes hair-bundle displacement to exceed basilar-membrane displacement. The theory of critical-layer absorption still applies. The basilar-membrane tuning curves, phase behavior, and nonlinearity consequently parallel those in regime I.

In regime III, the ratchet mechanism leads to very different behavior. Sound evokes a low-frequency pressure wave that traverses the entire cochlea, evoking only a small basilar-membrane displacement throughout, for the resonant frequency of the basilar-membrane is higher everywhere (Fig. 3B). At the characteristic place, however, the hair bundles exhibit an independent resonance amplified through active hair-bundle motility and facilitated by the unidirectional coupling provided by electromotility. The hair-bundle displacement there can exceed the basilar-membrane response by orders of magnitude (Fig. 3C). In further contrast to the theory of critical-layer absorption, the traveling wave does not slow and its wavelength does not vanish at the characteristic place, for the basilar-membrane does not resonate. This becomes apparent in the behavior of the traveling wave’s phase, whose slope remains low and nearly constant across the characteristic place (Fig. 3D). The phase behavior also confirms that the wave reaches the helicotrema. Because amplification through the ratchet mechanism does not act on the basilar membrane, the latter exhibits approximately linear behavior (Fig. 3C). The pressure wave is therefore unaffected by the active process. Hair-bundle displacement is amplified and exhibits a moderate compressive nonlinearity (Fig. 3C).

Threshold Tuning Curves of Auditory-Nerve Fibers. Strong support for the proposed model comes from studies of threshold tuning curves for auditory-nerve fibers (7–9). Recent measurements have shown characteristic shape changes occurring around the crossover frequencies f_H and f_L (9). To compare these data to our model, we have computed tuning curves and—despite their complexity and the simplicity of our model—found striking agreement with the measurements (Fig. 4). High-frequency fibers, those tuned above f_H , display the strongly asymmetric shape characteristic of critical-layer absorption. Moreover, they fall onto a universal curve when the frequency and threshold are measured relative to the values at the characteristic frequency. This accords with experimental findings and the scaling symmetry of the peaked traveling-wave mechanism (4, 5, 9). For intermediate

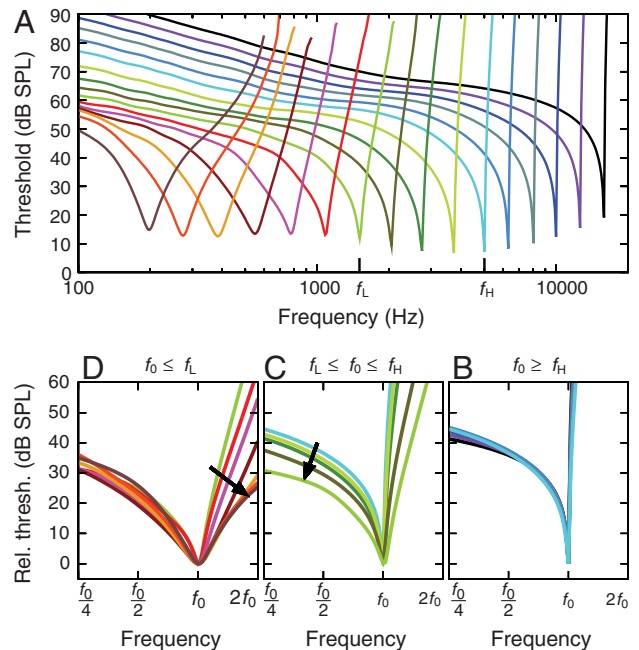


Fig. 4. Threshold tuning curves of auditory-nerve fibers. (A) The tuning curve at each position along the cochlea has a characteristic frequency f_0 corresponding to the resonant frequency of the hair-bundle complex. When tuning curves are rescaled such that the frequency is measured in octaves relative to the characteristic frequency f_0 and the threshold is measured relative to that at the characteristic frequency, characteristic shape changes are seen to occur between curves of different characteristic frequencies. (B) Tuning curves for high characteristic frequencies, above f_H , fall onto a universal curve that exhibits the strongly asymmetric form and high-frequency cutoff characteristic of a peaked traveling wave. (C) As the characteristic frequency declines from f_H to f_L , the left limb falls (Arrow), indicating the emerging influence of electromotility and the ratchet mechanism. (D) As the characteristic frequency diminishes below f_L , the right limb falls steeply (Arrow), pointing to the breakdown of the peaked-wave mechanism and the dominance of ratchet amplification.

characteristic frequencies between f_L and f_H the scaling law is violated as the ratchet mechanism starts to influence the micromechanics, causing the left limb of the threshold tuning curves to fall as the characteristic frequency decreases (9). A second violation of the scaling arises for characteristic frequencies below f_L , for which the right limb falls. Also observed experimentally (9), this second violation of scaling indicates a breakdown of critical-layer absorption that predicts a steep increase for threshold tuning curves at frequencies above the characteristic frequency.

Discussion

The classical theory of cochlear mechanics assumes that the basilar membrane resonates at a characteristic place for each frequency in the auditory range. The resulting mechanism of critical-layer absorption is characterized by (i) strongly asymmetric, scale-invariant tuning curves with steep, high-frequency cutoffs, (ii) an increasing slope of the traveling wave’s phase upon approaching the characteristic place, (iii) basilar-membrane displacement similar to hair-bundle displacement, and (iv) pronounced compressive nonlinearity at the characteristic frequency. This picture is consistent with diverse experimental findings from the cochlear base, including direct measurements of basilar-membrane motion and studies of threshold tuning curves for auditory-nerve fibers. However, as summarized previously, experimental results from the cochlea’s apex are in qualitative disagreement with the first, second, and fourth characteristics.

Here we have proposed a concept for low-frequency hearing that employs a ratchet mechanism involving the interplay of

active hair-bundle motility and electromotility. Whereas sound-evoked forces displace the basilar membrane and the hair bundles, the active forces within the hair bundle can decouple from the basilar membrane through appropriate elongation and contraction of OHCs. This mechanism allows hair bundles to resonate independently of the basilar membrane and thus explains how the ear's hearing range can extend to values well below the resonant frequency of the basilar membrane. We have shown that amplification by the ratchet mechanism leads to characteristic behavior that is distinct from that associated with critical-layer absorption: it exhibits (*i'*) approximately symmetric tuning curves that display no sharp, high-frequency cutoff, (*ii'*) a constant slope of the traveling wave's phase across the characteristic place, (*iii'*) tuned hair-bundle displacement that exceeds the untuned basilar-membrane displacement by orders of magnitude at resonance, and (*iv'*) approximately linear basilar-membrane displacement and only moderate compressive nonlinearity in hair-bundle motion at the characteristic frequency.

Comparison to Experimental Results. The proposed model yields the classical theory of critical-layer absorption for high-frequency sounds (Figs. 1*B*, 3, and 4*A*). Experiments in the base have confirmed (*i*) the asymmetric form of tuning curves (13, 14), (*ii*) the increasing slope of the phase (36, 37), and (*iv*) the strong compressive nonlinearity (13, 14, 37). Owing to difficulties in accessing the motion of the organ of Corti at the base, the relation (*iii*) of hair-bundle motion to basilar-membrane motion has not yet been measured there.

Because the membrane time constant of OHCs is expected to limit the operation of electromotility to low frequencies (33, 34), we have chosen in our model to consider only active hair-bundle motility in the basal part of the cochlea. However, studies in prestin knockout mice suggest that electromotility is necessary for high-frequency amplification (38, 39). Electromotility near the base may adjust the operating point of the hair-bundle motor on a time scale slower than the period of oscillation or provide a fast electrical signal in the form of membrane-potential change upon mechanical stimulation.

Our theory accounts for a number of unexplained results from the cochlear apex. Experiments have measured (*i'*) approximately symmetric tuning curves without sharp, high-frequency cutoffs of tectorial-membrane and therefore hair-bundle displacement (10, 11, 15). In our model, the approximate symmetry arises naturally, for it reflects the resonance of the hair bundles, reticular lamina, and tectorial membrane independently of resonance by the basilar membrane (Figs. 1*C* and 3*C* and *D*). Experiments have consistently reported (*ii'*) a constant phase slope across the characteristic place (10, 11, 40). This feature emerges in our model (Fig. 3*D*) because the basilar membrane is untuned at low frequencies and the pressure wave therefore reaches the helicotrema. There remains a controversy about the existence of nonlinearity at the cochlear apex (12): Although some investigators have found no nonlinearities (15), others have reported a small compressive (10, 16) or expansive nonlinearity (11). However, all agree about (*iv'*) the absence of a strong compressive nonlinearity.

The cochlear apex allows observation of different parts of the organ of Corti—such as the tectorial membrane and Hensen's cells as well as the basilar membrane—and thus permits tests of characteristic (*iii'*) in our model. Experiments on a temporal-bone preparation (15, 41) have shown that vibrations of the reticular lamina and tectorial membrane are orders of magnitude larger than the basilar-membrane displacement at the characteristic place. These findings are in perfect agreement with our theory. However, the *in vitro* experiments have been criticized for an uncertainty about which constituents of the organ of Corti were measured (12) and are in contradiction with studies using other

preparations (10, 16). Definite conclusions must, therefore, be postponed until further experimental results become available.

Perhaps the most reliable comparison of our model to experimental measurements comes through tuning curves of auditory-nerve fibers that yield information about cochlear mechanics that is least disturbed by experimental intervention and represent responses from the whole length of the cochlea. These tuning curves exhibit progressive shape changes from (*i*) the strongly asymmetric form typical of high-frequency fibers to (*i'*) the nearly symmetric form found for low-frequency fibers (7–9). Our modeled responses of tuning curves of auditory-nerve fibers exhibit the same shape changes (Fig. 4), thus providing a conceptual explanation for the three distinct cochlear regimes.

Experimentally Testable Predictions. Our theory yields experimentally testable predictions based on the characteristics (*i'*)–(*iv'*). As discussed above, (*i'*) the approximately symmetric tuning curves as well as (*ii'*) the constant phase slope have already been observed. In contrast, characteristic (*iii'*) that predicts hair-bundle motion that exceeds the basilar-membrane motion at the characteristic frequency by orders of magnitude remains controversial. Future experiments are therefore required to test this prediction. Such studies should also determine whether, as our theory implies, the complex formed by the hair bundles, reticular lamina, and tectorial membrane exhibits a resonance independent of the basilar membrane, whose response is untuned. A further test is feasible through experimental studies of the nonlinear behavior at the apex, for which our theory predicts (*iv'*) a moderate compressive nonlinearity in hair-bundle motion and therefore in tectorial-membrane motion, but no more than a weak nonlinearity in the basilar membrane's response. Finally, and beyond the theory presented in this article, the ratchet mechanism should allow for traveling waves in the tectorial membrane (42) that are unidirectionally coupled to basilar membrane waves.

The Ratchet Principle. The ratchet mechanism constitutes a general design principle for mechanical amplification whereby the output does not feed back onto the input. In this way, it represents a mechanical analogue of the operational amplifier from electrical engineering (28). Although the current technology of signal detectors such as microphones relies on electrical amplification, mechanical amplification could improve signal-to-noise ratios and thereby greatly advance sensitivity and detection of weak signals. The ratchet mechanism thus opens a path for implementing controlled mechanical amplification in engineering applications.

Materials and Methods

Hydrodynamics. By combining continuity equations and fluid-momentum equations, we describe the cochlea's hydrodynamics with the partial differential equation

$$\rho \partial_r^2 X_{BM} + \Lambda \partial_r X_{BM} = \frac{1}{2L^2} \partial_r (h \partial_r p). \quad [4]$$

Here ρ denotes the density of liquid in the cochlea, L the length of the cochlea, h the height of the scalae, and p and X_{BM} , respectively, the pressure across and the displacement of the basilar-membrane at position r and time t . The coefficient Λ accounts for friction due to fluid motion. Position is measured in units of the cochlear length, such that $r = 0$ corresponds to the basal and $r = 1$ to the apical end. The pressure translates into an external force acting on the basilar membrane and yields a displacement that we compute employing the model of Fig. 2*B* (SI Text). Temporal Fourier transformation yields an ordinary differential equation that we solve numerically with the shooting method in Mathematica 7 (Wolfram Research). We apply two boundary conditions. First, $p = p_0$ at $r = 0$: a sound-evoked pressure p_0 acts at the stapes. And second, $p = 0$ at $r = 1$: because the two scalae communicate at the helicotrema, the pressure difference between them vanishes at the apical end of the cochlea.

Tuning Curves of Auditory-Nerve Fibers. Tuning curves of auditory-nerve fibers are computed by assuming that the hearing threshold corresponds to a root-mean-square deflection of the hair bundles of inner hair cells by 0.3 nm. These bundles are thought to be coupled by fluid motion to the shearing between the reticular lamina and tectorial membrane and thus to the displacement of the hair bundles of OHCs (*SI Text*).

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Supporting Information

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SI Text

Here we further describe our modeling approach. We provide a detailed description of the two-mass model for the organ of Corti and discuss the energy balance. We then turn to the hydrodynamics of the cochlea. Because our goal is to demonstrate a principle rather than to describe the cochlea in full detail, we focus on a basic, one-dimensional model. We conclude with a model for the regulation of the OHCs' membrane potential, which demonstrates how the critical mechanomotility coefficient α_* can result.

Micromechanics of the Organ of Corti. To describe the micromechanics of the organ of Corti we employ a model with two degrees of freedom: the motion of the basilar membrane and the motion of the hair-bundle complex (Fig. 2B). The impedance Z_{BM} of the basilar membrane results from a mass m_{BM} , viscous damping λ_{BM} , and stiffness K_{BM} . Similarly, the impedance of the complex formed by the hair bundles, reticular lamina, and tectorial membrane involves mass m_{TM} , viscous damping λ_{HB} , and stiffness K_{HB} .

The motions of the basilar membrane and of the hair-bundle complex are coupled through the impedance Z_C of the organ of Corti, which possesses viscous and elastic contributions λ_C and K_C . Further coupling arises through the combined OHCs and Deiters' cells (Fig. S1). An OHC can be described as a piezoelectric element (1). Its length change δL depends both on a change δV in the membrane potential as well as on the applied force F . Considering oscillatory motions at angular frequency $\omega = 2\pi f$ we can write $\delta L = \delta L e^{i\omega t} + c.c.$, $\delta V = \delta V e^{i\omega t} + c.c.$, and $F = F e^{i\omega t} + c.c.$ in which $c.c.$ denotes the complex conjugate. The length change then follows as

$$\widetilde{\delta L} = -c\widetilde{\delta V} + c_2\widetilde{F} \quad [S1]$$

with coefficients c, c_2 . We define $\widetilde{X}_{EE} = -c\widetilde{\delta V}$ as the part of the length change that results from voltage changes alone. The length change $c_2\widetilde{F}$ results from the OHC's viscoelasticity $Z_{OHC} = -i\omega^{-1}c_2^{-1}$ in series with X_{EE} (Fig. S1). It lies in series with the Deiters' cell which can be described as another viscoelastic element Z_{DC} . The two viscoelastic elements Z_{OHC} and Z_{DC} in series combine into the viscoelastic element

$$Z_D = (Z_{OHC}^{-1} + Z_{DC}^{-1})^{-1} \quad [S2]$$

such that the schematic of Fig. 2B results. We denote by K_D the elastic component and by λ_D the viscous part of the impedance Z_D . The equations of motion for hair-bundle and basilar-membrane displacement, X_{HB} and X_{BM} , respectively, then read

$$\begin{aligned} m_{TM}\partial_t^2 X_{HB} + \lambda_{HB}\partial_t X_{HB} + K_{HB}X_{HB} \\ + (K_D + \lambda_D\partial_t)(X_{HB} - X_{EE} - X_{BM}) \\ + (K_C + \lambda_C\partial_t)(X_{HB} - X_{BM}) \\ = F_{int}, \end{aligned} \quad [S3]$$

$$\begin{aligned} m_{BM}\partial_t^2 X_{BM} + \lambda_{BM}\partial_t X_{BM} + K_{BM}X_{BM} \\ - (K_D + \lambda_D\partial_t)(X_{HB} - X_{EE} - X_{BM}) \\ - (K_C + \lambda_C\partial_t)(X_{HB} - X_{BM}) \\ = F_{ext}. \end{aligned} \quad [S4]$$

A sound stimulus at angular frequency ω provides an external force F_{ext} on the basilar membrane whose dependence on time t may be written as $F_{ext}(t) = \widetilde{F}_{ext}e^{i\omega t} + c.c.$ with the Fourier coefficient \widetilde{F}_{ext} . The evoked oscillations of the basilar membrane, X_{BM} , and of the hair-bundle complex, X_{HB} , occur dominantly at the same frequency f ; additional frequencies may arise from nonlinear effects owing to internal forces in the hair bundle. We consider the corresponding Fourier coefficients \widetilde{X}_{BM} and \widetilde{X}_{HB} . The electrically evoked length change \widetilde{X}_{EE} of OHCs depends linearly on the hair-bundle displacement, for the membrane-potential change depends linearly on hair-bundle motion (see Supplementary Section on the electrical regulation of the OHC membrane potential): $\widetilde{X}_{EE} = -\alpha\widetilde{X}_{BM}$ with a complex, frequency-dependent mechanomotility coefficient α . Eq. S4 then yields Eqs. 1 and 2 with

$$Z_{HB} = i\omega m_{TM} + \lambda_{HB} - i\omega^{-1}K_{HB}, \quad [S5]$$

$$Z_{BM} = i\omega m_{BM} + \lambda_{BM} - i\omega^{-1}K_{BM}, \quad [S6]$$

$$Z_C = \lambda_C - i\omega^{-1}K_C, \quad [S7]$$

$$Z_D = \lambda_D - i\omega^{-1}K_D. \quad [S8]$$

The forces within the hair bundle depend on its displacement; their effect is to counter viscous damping as well as to influence the resonant frequency of the hair-bundle complex. We may decompose these forces into linear and nonlinear parts:

$$\widetilde{F}_{int} = i\omega Z_{HB}^{act}\widetilde{X}_{HB} + \widetilde{F}_{int}^{nonlin}(\widetilde{X}_{HB}). \quad [S9]$$

Here the linear term dominates the dynamics for small displacements ensuing from weak sound stimuli and describes the fully active scenario. The nonlinear term connects the active to the passive case that arises for strong sound stimuli.

At the critical value α_* , the displacements are given by Eq. 3, which can be rewritten using Eq. S9 as

$$\begin{aligned} \widetilde{X}_{HB} = \frac{1}{i\omega(Z_{HB} - Z_{HB}^{act})}\widetilde{F}_{int}^{nonlin}(\widetilde{X}_{HB}) \\ + \frac{Z_D + Z_C}{i\omega(Z_{HB} - Z_{HB}^{act})(Z_{BM} + Z_D + Z_C)}\widetilde{F}_{ext}, \end{aligned} \quad [S10]$$

$$\widetilde{X}_{BM} = \frac{1}{i\omega(Z_{BM} + Z_D + Z_C)}\widetilde{F}_{ext}. \quad [S11]$$

Resonance in hair-bundle motion occurs at the frequency for which $Z_{HB} - Z_{HB}^{act} = 0$. Active hair-bundle motility makes this situation possible. As discussed in the main text, this resonance condition is independent of the properties of the basilar membrane.

Cochlear Hydrodynamics and Traveling Waves. We now incorporate the description for the micromechanics of the organ of Corti into a model for the whole cochlea. In the simplest form, the cochlea is considered as one-dimensional, exhibiting a slow pressure wave. Combining continuity equations and fluid-momentum

equations, this pressure wave obeys the partial differential equation

$$\rho \partial_t^2 X_{\text{BM}} + \Lambda \partial_t X_{\text{BM}} = \frac{1}{2L^2} \partial_r (h \partial_r p). \quad [\text{S12}]$$

Here ρ denotes the density of liquid in the cochlea, L the length of the cochlea, h the height of the scalae, and p and X_{BM} respectively the pressure across and the displacement of the basilar membrane at position r and time t . The coefficient Λ accounts for friction due to fluid motion. Position is measured in units of the cochlear length, such that $r = 0$ corresponds to the basal and $r = 1$ to the apical end. We apply two boundary conditions. First, $p = p_0$ at $r = 0$: a sound-evoked pressure p_0 acts at the stapes. And second, $p = 0$ at $r = 1$: because the two scalae communicate at the helicotrema, the pressure difference between them vanishes at the apical end of the cochlea.

Considering the Fourier coefficients of angular frequency ω , we obtain

$$-\omega^2 \rho \tilde{X}_{\text{BM}} + i\omega \Lambda \tilde{X}_{\text{BM}} = \frac{1}{2L^2} \partial_r (h \partial_r \tilde{p}). \quad [\text{S13}]$$

The basilar-membrane displacement \tilde{X}_{BM} and pressure \tilde{p} are connected through Eq. 1, with $\tilde{p} = \tilde{F}_{\text{ext}}/A_{\text{BM}}$. A_{BM} denotes the area of a transverse strip of the basilar membrane that has the width of one cell, about $8 \mu\text{m}$; all parameters employed in the two-mass model refer to a transverse segment of the cochlear partition of that width. We solve Eq. S13 numerically with the shooting method in Mathematica 7 (Wolfram Research).

The height h of the scalae varies from about 1 mm at the base to about $200 \mu\text{m}$ at the apex. Measurements of the fluid pressure in the basal region indicate, however, that the penetration depth of the wave is significantly smaller (2). In our one-dimensional simulations, we therefore use an effective height of $100 \mu\text{m}$.

Concerning the map of characteristic frequencies $f_0(r)$ along the cochlea, we consider the chinchilla's hearing range from a maximal frequency of about 30 kHz at the extreme base to a minimum of about 50 Hz at the extreme apex (3). We assume that the resonant frequency of the basilar membrane cannot cover this whole range and is instead given by

$$f_{\text{BM}}(r) = \sqrt{f_{\text{BM,apex}}^2 + [1 - (f_{\text{BM,apex}}/f_{\text{max}})^2]f_0(r)^2}. \quad [\text{S14}]$$

Here $f_{\text{max}} = 30 \text{ kHz}$ denotes the highest characteristic frequency and $f_{\text{BM,apex}} = 1 \text{ kHz}$ the lowest resonant frequency that the basilar membrane exhibits. f_{BM} coincides with f_0 for high frequencies but deviates for frequencies below f_{L} and approaches $f_{\text{BM,apex}}$ in the apical region of the cochlea (Fig. 3B).

The resonant frequency f_{BM} of the basilar membrane is set by its mass and stiffness according to $f_{\text{BM}} = (2\pi)^{-1} \sqrt{K_{\text{BM}}/m_{\text{BM}}}$. We consider K_{BM} to vary proportional to f_{BM} and m_{BM} to vary inversely: $K_{\text{BM}}(r) = K_{\text{BM}}^{\text{max}} f_{\text{BM}}(r)/f_{\text{max}}$ and $m_{\text{BM}}(r) = K_{\text{BM}}(r)/[2\pi f_{\text{BM}}(r)]^2$, with $K_{\text{BM}}^{\text{max}} = 2 \text{ N}\cdot\text{m}^{-1}$. The mass m_{TM} of the tectorial membrane is assumed to follow the same spatial variation as the basilar membrane masses, albeit with a smaller value: $m_{\text{TM}} = m_{\text{BM}}/5$. The increase in the basilar-membrane and tectorial-membrane masses from base to apex presumably reflects the increasing size of the organ of Corti towards the apex.

For the variation of the mechanomotility coefficient α along the cochlea, and its dependence on the stimulus frequency f , we make the ansatz

$$\alpha(r, f) = \frac{f_c^4}{f_c^4 + f_0(r)^4} \times \frac{[\delta f(r) f_0]^2}{[f^2 - f_0(r)^2]^2 + [\delta f f_0(r)]^2} \times \alpha_*. \quad [\text{S15}]$$

The first factor accounts for the high-frequency cutoff above which the membrane potential, and thus electromotility, can

no longer follow hair-bundle displacement on a cycle-by-cycle basis; we use $f_c = 4 \text{ kHz}$. To yield the ratchet mechanism at lower frequencies, we choose α to be α_* at the natural frequency. The second factor describes how the response changes when the frequency f of stimulation deviates from the natural frequency f_0 ; we assume that α is close to α_* as long as both frequencies are similar, but otherwise declines in magnitude. The width of the corresponding curve is determined by the parameter δ , which we set at $\delta = 2$. The mechanomotility coefficient $\alpha(r, f = f_0)$ is shown in Fig. 3A.

The linear part of the active hair-bundle force counters viscous damping and provides a vanishing impedance at the natural frequency. Using Eqs. 1, 2, and S9, this translates into

$$Z_{\text{HB}} - Z_{\text{HB}}^{\text{act}}|_{f=f_0} = - \frac{Z_{\text{BM}}[(1 + \alpha)Z_{\text{D}} + Z_{\text{C}}]}{Z_{\text{BM}} + Z_{\text{C}} + Z_{\text{D}}}|_{f=f_0}. \quad [\text{S16}]$$

We assume that the imaginary part of Z_{HB} is tuned to the natural frequency f_0 such that the active contribution possesses only a real part corresponding to negative damping, $Z_{\text{HB}}^{\text{act}} \equiv \lambda_{\text{HB}}^{\text{act}}$. Note that in the ratchet mechanism, when $\alpha = \alpha_*$, the right-hand side of Eq. S16 vanishes, so active hair-bundle forces need to overcome only damping associated with hair-bundle motion.

When α and active hair-bundle forces obey Eqs. S15 and S16 and nonlinearities from the hair-bundle forces are ignored, the hair-bundle displacement diverges at resonance. In the actual cochlea, noise as well as nonlinearities supervene and lead to a large but finite displacement. In our simulations, we incorporate this effect by considering values for α and $Z_{\text{HB}}^{\text{act}}$ that deviate by 1% from their ideal values given by Eqs. S15 and S16.

Tuning Curves of Auditory-Nerve Fibers Tuning curves of auditory-nerve fibers are computed by assuming that the hearing threshold corresponds to a root-mean-square deflection of the hair bundles of IHCs by 0.3 nm. These bundles are thought to be coupled by fluid motion to the shearing between the reticular lamina and tectorial membrane, and thus to the displacement of the hair bundles of OHCs. Assuming that the displacement X_{IHB} of the hair bundles of inner hair cells is dependent on friction λ_{IHB} and elasticity K_{IHB} , it couples to the OHCs' bundle motion through the relation

$$\lambda_{\text{IHB}} \partial_t (X_{\text{IHB}} - X_{\text{HB}}) + K_{\text{IHB}} X_{\text{IHB}} = 0. \quad [\text{S17}]$$

Upon Fourier transformation, we obtain

$$\tilde{X}_{\text{IHB}} = \frac{i\omega \lambda_{\text{IHB}}}{i\omega \lambda_{\text{IHB}} + K_{\text{IHB}}} \tilde{X}_{\text{HB}}. \quad [\text{S18}]$$

For high frequencies, viscous coupling is strong and the hair-bundle displacements of IHCs and OHCs coincide. For low frequencies, the coupling decreases and the hair-bundle motion of IHCs remains smaller than that of OHCs. This effect underlies the rising thresholds at low frequencies seen on the left limbs of high-frequency tuning curves (Figure 5) as well as in experimental measurements.

The value of 0 dB SPL is defined by a root-mean-square sound-pressure stimulus of $20 \mu\text{Pa}$. Because this external pressure is enhanced by the middle ear before acting on the stapes, we assume an increase by a factor of 20 (26 dB) (9).

If they have not been detailed in the text above, the parameters of the two-mass model as well as the one-dimensional descriptions of the cochlea and hair-bundle dynamics of IHCs are given in Table S1.

Electrical Regulation of the OHC Membrane Potential. Deflection X_{HB} of the OHCs hair bundles evokes a change $\delta V \equiv V - V_0$ in the membrane potential from its resting value V_0 , which leads to a length change $X_{\text{EE}} = -c\delta V$ (see Eq. S1 and below) with a

coefficient c of about $20 \mu\text{m} \cdot \text{V}^{-1}$ (1). The ratchet mechanism relies on a critical value $\alpha_* = -1 - Z_C/Z_D$ of the ratio $\alpha = -X_{EE}/X_{HB}$ between hair-bundle displacement and cell length change. While its exact value depends on the coupling impedances Z_C and Z_D , the real part of α_* is negative. A typical situation $Z_C = Z_D$ leads to the critical value $\alpha_* = -2$. Here we show how such a value of α_* can emerge within a basic model for the regulation of the OHCs membrane potential. The model relies on voltage-regulated outward K^+ channels (10) that counteract the current through the membrane capacitance, as well as a voltage-regulated inward sodium current (11, 12) that can lead to hyperpolarization upon positive hair-bundle deflection.

The apical surface of an OHC, including the hair bundle, is bathed in endolymph at a potential V_E of about 80 mV (Fig. S2). The basolateral part is surrounded by perilymph at ground potential V_P . The electrical behavior can be represented by a simple circuit described by an equation for current conservation:

$$I_{R_a} = I_{R_b} + I_{C_b}. \quad [\text{S19}]$$

Here I_{R_a} denotes the current through the resistance R_a and I_{R_b} and I_{C_b} denote the currents through R_b and C_b , respectively. We assume that R_a results from the conductivity g_a of the mechanotransduction channels in the hair bundle; the current is mainly carried by K^+ ions (13). The conductance of the basolateral cell membrane stems from voltage-regulated K^+ channels (conductance g_b^K) as well as voltage-regulated Na^+ channels (11, 12) (conductance g_b^{Na}). We obtain

$$g_a(V_E - V) = C_b \partial_t V + g_b^K(V - V_P) - g_b^{\text{Na}}(V_{\text{Na}} - V) \quad [\text{S20}]$$

in which V_{Na} denotes the Na^+ reversal potential and C_b the cell membrane's capacitance. The potential V_0 inside the cell at vanishing hair-bundle displacement $X_{\text{HB}} = 0$ follows as

$$V_0 = \frac{g_a^{(0)}V_E + g_b^{\text{Na}(0)}V_{\text{Na}} + g_b^{\text{K}(0)}V_P}{g_a^{(0)} + g_b^{\text{Na}(0)} + g_b^{\text{K}(0)}} \quad [\text{S21}]$$

in which the superscript (0) denotes the respective conductances at vanishing hair-bundle displacement.

Hair-bundle displacement X_{HB} changes the conductance g_a of the cell's apical membrane. To linear order we obtain

$$g_a = g_a^{(0)} + \left. \frac{\partial g_a}{\partial X_{\text{HB}}} \right|_{(0)} X_{\text{HB}}. \quad [\text{S22}]$$

The altered apical conductance evokes a change $\delta V = V - V_0$ in the membrane potential which influences the basal K^+ and Na^+

conductances. The K^+ channels are voltage-regulated at an activation time τ . The dynamics of the change $\delta g_b^K = g_b^K - g_b^{\text{K}(0)}$ can be described by

$$\tau \partial_t \delta g_b^K = \left. \frac{\partial g_b^K}{\partial V} \right|_{(0)} \delta V - \delta g_b^K \quad [\text{S23}]$$

which results in

$$\tilde{\delta g_b^K} = \frac{1}{1 + i\omega\tau} \left. \frac{\partial g_b^K}{\partial V} \right|_{(0)} \tilde{\delta V} \quad [\text{S24}]$$

for oscillatory stimuli $X_{\text{HB}} = \tilde{X}_{\text{HB}} e^{i\omega t} + c.c.$ at angular frequency ω . The Na^+ channels are rapidly activated; to linear order their conductance follows as

$$g_b^{\text{Na}} = g_b^{\text{Na}(0)} + \left. \frac{\partial g_b^{\text{Na}}}{\partial V} \right|_{(0)} \delta V. \quad [\text{S25}]$$

Linearization of Eq. S20 yields

$$\begin{aligned} [i\omega C_b + g_a + g_b^K + g_b^{\text{Na}} + \frac{1 - i\omega\tau}{1 + \omega^2\tau^2} \left. \frac{\partial g_b^K}{\partial V} \right|_{(0)} (V - V_P) \\ - \left. \frac{\partial g_b^{\text{Na}}}{\partial V} \right|_{(0)} (V_{\text{Na}} - V)] \tilde{\delta V} \\ = \left. \frac{\partial g_a}{\partial X_{\text{HB}}} \right|_{(0)} \tilde{X}_{\text{HB}}. \end{aligned} \quad [\text{S26}]$$

The mechanomotility coefficient α follows as $\alpha = c \tilde{\delta V} / \tilde{X}_{\text{HB}}$. From Eq. S26 we note that delayed voltage-regulated K^+ channels can reduce the effect of the membrane capacitance. A large enough voltage dependence of the inward Na^+ channels can then yield hyperpolarization of the cell upon positive deflection of the hair bundle, as requested for the critical value α_* .

Fig. S3 shows the resulting magnitude and phase of α for typical parameters for apical OHCs (Table S2) in dependence on the frequency of hair-bundle stimulation. The values $\partial g_b^K / \partial V = 1.36 \mu\text{S} \cdot \text{V}^{-1}$ and $\partial g_b^{\text{Na}} / \partial V = 1.85 \mu\text{S} \cdot \text{V}^{-1}$ have been chosen to yield the critical value $\alpha_* = -2$ at the frequency $f_0 = 200$ Hz and lie in the range of measured values (10, 11).

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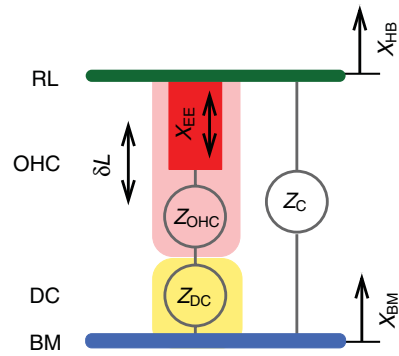


Fig. S1. Coupling between the reticular lamina (RL) and the basilar membrane (BM). The coupling arises from OHCs in series with Deiters' cells (DC) and an impedance Z_C from the remaining organ of Corti. See the text for a description of the coupled OHCs and Deiters' cells.

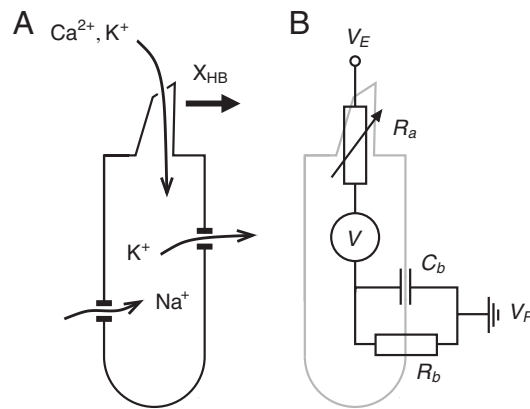


Fig. S2. Regulation of the OHC membrane potential. (A) Hair-bundle deflection opens ion channels; K^+ and Ca^{2+} flow into the cell. The cell membrane contains various additional ion channels that are regulated by the membrane potential and the Ca^{2+} concentration. In our model, we consider voltage-regulated outward K^+ channels and inward Na^+ channels. (B) An equivalent circuit of an OHC with a variable resistance R_a controlled by hair-bundle displacement, a membrane capacitance C_b , and a basolateral membrane conductance R_b .

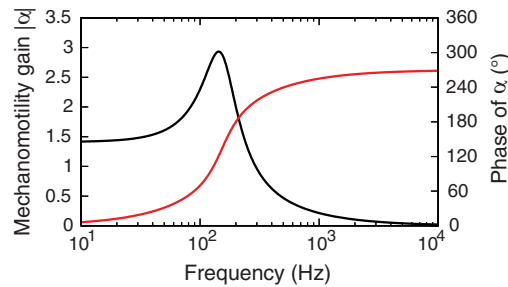


Fig. S3. The modeled mechanomotility coefficient α . We show the dependence of the magnitude (Black) and phase (Red) of α on the sound frequency. For $f_0 = 200$ Hz the coefficient $\alpha = -2$ emerges, which represents a typical value for the critical α_* . The magnitude of α is strongly attenuated for higher frequencies. By varying the voltage dependence of the basal K^+ and Na^+ channels the critical value α_* can emerge at different frequencies f_0 below a few kilohertz.

Table S1. Parameters used in the cochlear model

Parameter	Description	Value	Reference
L	length of the cochlea	20 mm	[4]
Λ	fluid friction	$5 \times 10^5 \text{ N} \cdot \text{s} \cdot \text{m}^{-4}$	
$w_{\text{BM}}(r)$	width of the basilar membrane	$(50 + 200r) \mu\text{m}$	[5]
$A_{\text{BM}}(r)$	area of the basilar-membrane strip	$w_{\text{BM}}(r) \times 8 \mu\text{m}$	
λ_{HB}	friction of the hair-bundle complex	$1 \mu\text{N} \cdot \text{s} \cdot \text{m}^{-1}$	[6]
K_{IHB}	stiffness of the hair bundles of IHCs	$2 \text{ mN} \cdot \text{m}^{-1}$	[7]
λ_{IHB}	friction of the hair bundles of IHCs	$1 \mu\text{N} \cdot \text{s} \cdot \text{m}^{-1}$	[6]
$\lambda_{\text{BM}}(r)$	friction of the basilar membrane	$0.03 \times w_{\text{BM}}(r) \text{ N} \cdot \text{s} \cdot \text{m}^{-2}$	
K_{C}	stiffness of the organ of Corti	$10 \text{ mN} \cdot \text{m}^{-1}$	
λ_{C}	friction of the organ of Corti	$10 \mu\text{N} \cdot \text{s} \cdot \text{m}^{-1}$	
K_{D}	stiffness of OHCs and Deiters' cells	$20 \text{ mN} \cdot \text{m}^{-1}$	[8]
λ_{D}	friction of OHCs and Deiters' cells	$10 \mu\text{N} \cdot \text{s} \cdot \text{m}^{-1}$	

Table S2. Parameters for modeling the regulation of the OHCs membrane potential

Parameter	Description	Value	Reference
V_E	potential of the endolymph	80 mV	[14]
V_P	potential of the perilymph	-100 mV	[14]
V_{Na}	Na^+ reversal potential	22 mV	[11]
$g_a^{\text{K}^+}$	apical conductivity of K^+ channels	5 nS	[15]
$g_b^{\text{K}^+}$	basolateral K^+ conductivity	10 nS	[10]
C_b	membrane capacitance	40 pF	[10]
$\partial g_a^{\text{K}^+} / \partial X_{\text{HB}}$	dependence of apical K^+ conductivity on X_{HB}	$5 \text{ mS} \cdot \text{m}^{-1}$	[16]
τ	activation time of voltage-regulated K^+ channels	1 ms	[10]



Movie S1 Animated illustration of fundamental modes of motion by the organ of Corti. The passive motion is modest, and the hair-bundle movement is similar to that of the basilar membrane. Active hair-bundle motility greatly enhances the motion of both the hair bundles and the basilar membrane. In the ratchet mechanism, only hair-bundle motion is amplified; the basilar membrane moves as in the passive case.

[Movie S1 \(MOV\)](#)